

The Chloroplast Electron Transport Chain

Peter Gawthrop (peter.gawthrop@unimelb.edu.au)

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Note: this is the Chloroplast.ipynb notebook. The PDF version "The Chloroplast Electron Transport Chain" is available [here](#).

This is a work in progress and needs more explanatory notes.

```
In [1]: ## Some useful imports
import BondGraphTools as bgt
import numpy as np
import sympy as sp
import matplotlib.pyplot as plt

## Stoichiometric analysis
import stoich as st

## SVG bg representation conversion
import svgBondGraph as sbg

## Modular bond graphs
import modularBondGraph as mbg

## Display (eg disp.SVG(), disp.
import IPython.display as disp

## Data
import phiData
import redoxData
```

1 Introduction

Photosynthesis within plant chloroplasts is the basis of life on earth (Blankenship, 2015), (Nicholls and Ferguson, 2013).

Like the mitochondrion, the chloroplast has a membrane separating an inner space (lumen) from an outer space (stroma). In the chloroplast, the lumen gains protons and is called the p-space, the stroma loses protons and is called the n-space. Thus geometrically, the lumen corresponds to the mitochondrial matrix and the stroma to the mitochondrial intermembrane space; but electrically the p-space is inside and the n-space outside - the reverse of the mitochondrial situation.

The chloroplast electron transport chain has 4 complexes.

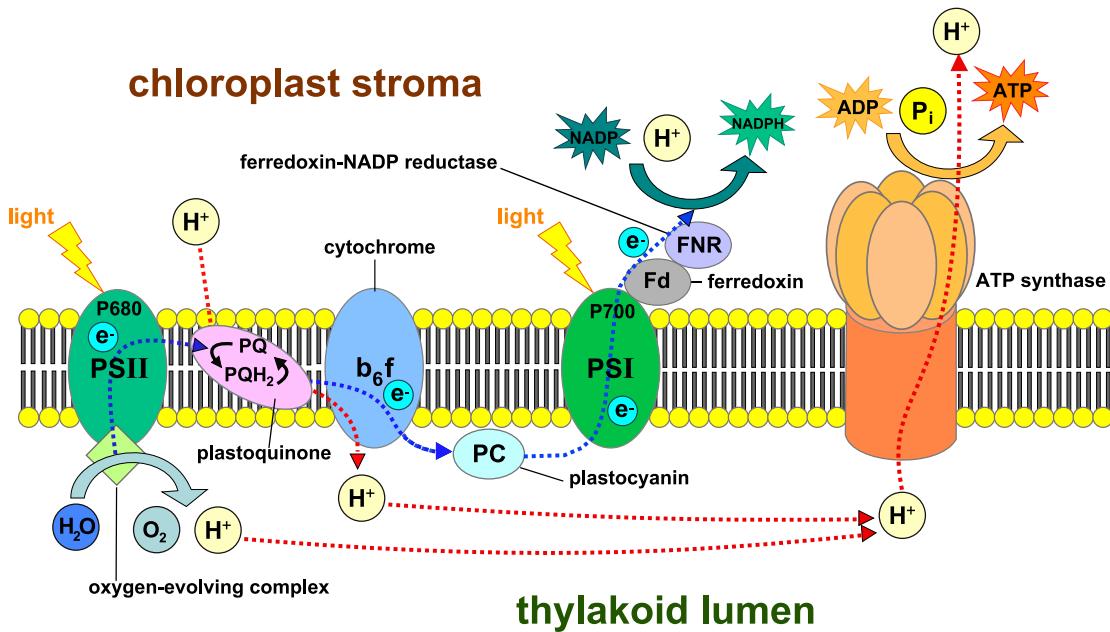
1. Photosystem II (PII) which absorbs photons at 680nm and splits water releasing protons into the p-space and passing electrons to the plastoquinone(PQ)/plastoquione(PQH₂) couple which absorbs protons from the n-space.
2. Cytochrome bf (Cyt) which passes electrons to the plastoquine/plastoquinone couple which releases two protons into the p-space. Electrons are passed to the plastocyanine couple (PcOx/PcRed). Two protons are pumped across the membrane.
3. Photosystem I (PI) which absorbs photons at 700nm and transports electrons from the plastocyanine (PcRed/PcOx) couple to the ferredoxin (FdOx/FdRed) couple.

4. Feredoxin-NADP reductase which transfers electrons from the ferredoxin (FdRed/FdOx) couple to convert NADP to NADPH absorbing a proton from the n-space.

The following figure is: https://commons.wikimedia.org/wiki/File:Thylakoid_membrane_3.svg

```
In [2]: # https://commons.wikimedia.org/wiki/File:Thylakoid_membrane_3.svg
disp.SVG("Thylakoid_membrane_3.svg")
```

Out [2] :



2 Photon Energetics

$$\phi_{photon} = \frac{N_{av}hc}{F\lambda} \quad (1)$$

where N_{av} = Avogadro's number

h = Planck's constant

c = velocity of light

F = Faraday's constant

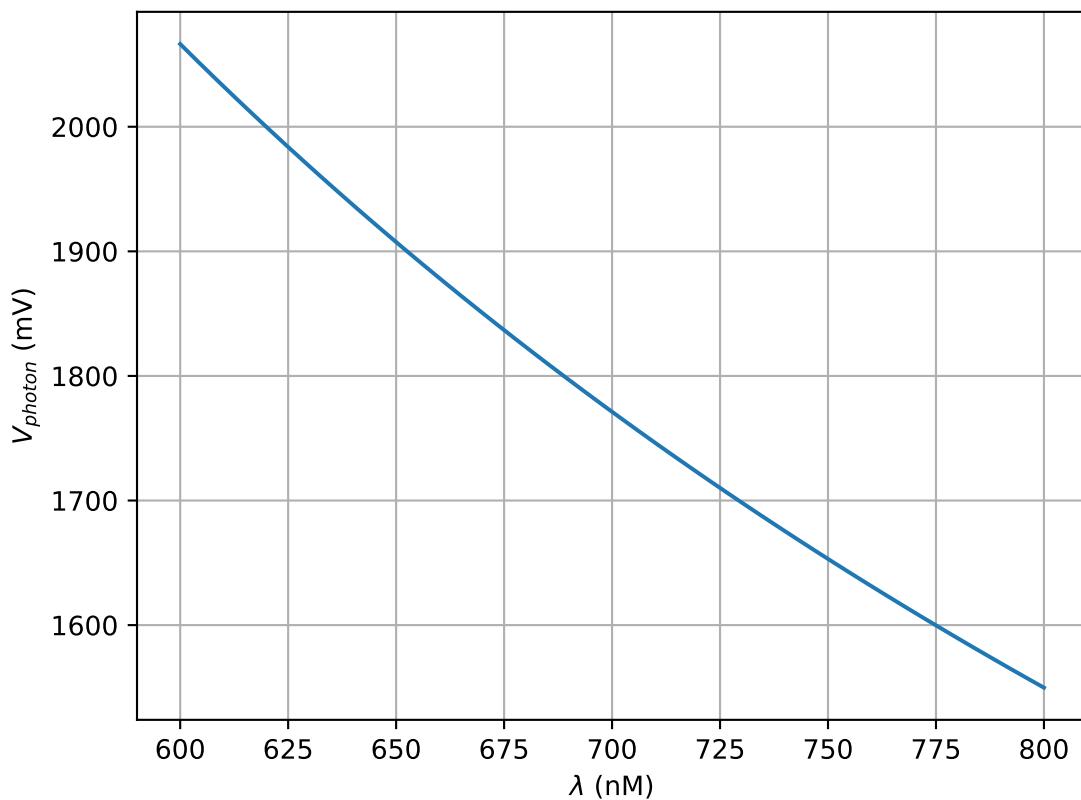
and λ = wavelength

For example:

$$\phi_{photon} = \begin{cases} 1.82V & \lambda = 680nm \\ 1.77V & \lambda = 700nm \end{cases} \quad (7)$$

In [3]: `disp.SVG('V_photon.svg')`

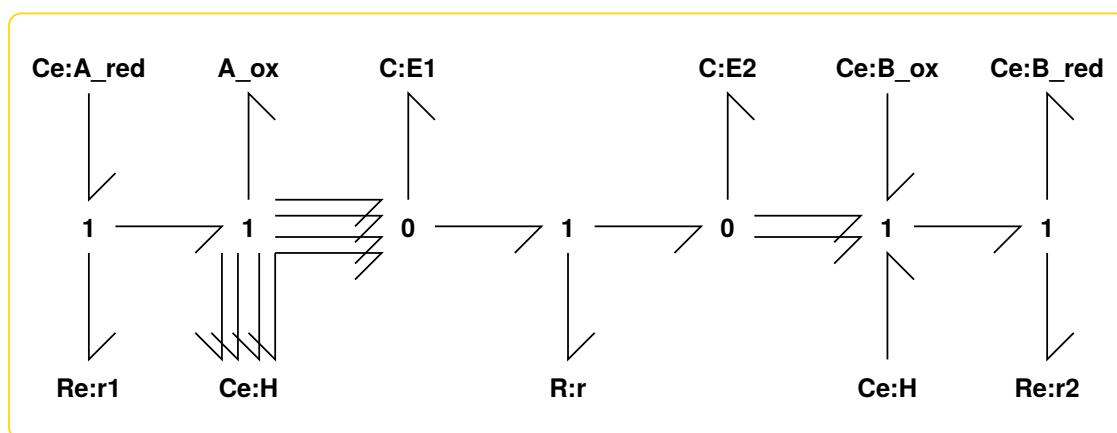
Out [3] :



3 Redox reactions

In [4]: `disp.SVG('Redox_abg.svg')`

Out [4] :



Redox reactions can be written as two half-reactions:



3.1 Redox potentials

Note that photon energies have been used for V_700 and V_680; this value is too large as the energy conversion is not direct. A model of this needs to be built. See, for example: ([Blankenship and Prince, 1985](#))

```
In [5]: import redoxData
      ## pH from BerTymStr 19.3
      pH_p = 4 # pH of p-space
      pH_n = pH_p + 3
      VpH = redoxData.VpH(pH_p - pH_n)
      V680 = redoxData.V_photon(wavelength=680)
      V700 = redoxData.V_photon(wavelength=700)
      print(VpH)
      print('V_pH =', int(1000*VpH), 'mV')
      print('V_680 =', int(1000*V680), 'mV')
      print('V_700 =', int(1000*V700), 'mV')
```

0.18462115653058278

```
V_pH = 184 mV
V_680 = 1823 mV
V_700 = 1771 mV
```

```
In [6]: ## Convert redox potentials to species phi
phi_redox = redoxData.phi()
phi_redox['Hn'] = redoxData.VpH(pH_n)
phi_redox['Hp'] = redoxData.VpH(pH_p)

## The correct photon potentials need sorting out
## The raw values used here are an overestimate. See BlaPri85
phi_redox['P680'] = V680
phi_redox['P700'] = V700
#phi_redox['P680'] = phi_redox['P680+']
#phi_redox['P700'] = phi_redox['P700+']

## The membrane potential is said by some to be zero. (check this)
phi_redox['dV'] = VpH
```

4 Bond graph description

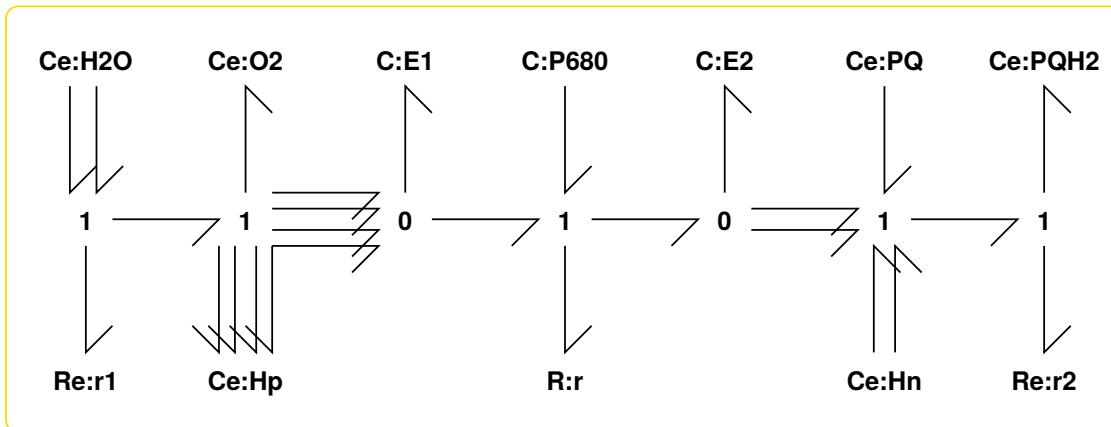
The four complexes are represented by svg graphics which are automatically converted into BondGraphTools format.

The stoichiometric toolbox is then used to generate the pathway-reduced equation for the complex.

4.1 Complex PII -- Photosystem II

In [7]: `disp.SVG('PII_abg.svg')`

Out [7] :



In [8]: `sbg.model('PII_abg.svg', convertR=True, convertCe=True, quiet=True)`
`import PII_abg`

```
In [9]: ## Stoichiometry
sPSII = st.stoich(PII_abg.model(), quiet=True)
chemostats = ['H2O', 'O2', 'PQ', 'PQH2', 'Hn', 'Hp', 'P680']
scPSII = st.statify(sPSII, chemostats=chemostats)
spPSII = st.path(sPSII, scPSII)
## All reactions
disp.Latex(st.sprintrl(sPSII, chemformula=True))
```

Out [9] :



In [10]: ## Pathway reaction
`disp.Latex(st.sprintrl(spPSII, chemformula=True))`

Out[10] :



```
In [11]: ## Compute net redox potential
RP_PII = (
    - redoxData.EpH('O2/2H2O', pH=pH_p)
    + V680
    + redoxData.EpH('PQ/PQH2', pH=pH_n)
)

print(redoxData.EpH('O2/2H2O', pH=pH_p))
print(redoxData.E('P680+/P680*'))
print(redoxData.E7('PQ/PQH2'))
print(redoxData.EpH('PQ/PQH2', pH=pH_n))
#print(RP_PII)
print('RP_PII =', int(1000*RP_PII), 'mV')

1.0006211565305827
0.8
0
0.0
RP_PII = 822 mV
```

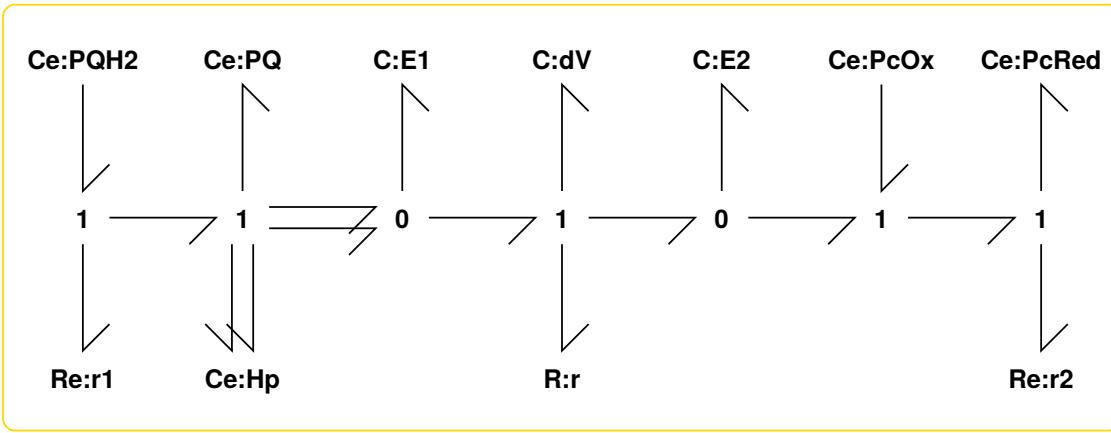
```
In [12]: ## Compute the reaction potential Phi
phi = phiData.phi_species(phi_redox, spPII['species'])
Phi_PII_ = -spPII['N'].T@phi
Phi_PII = Phi_PII_[0][0]
print('Phi_PII =', int(1000*Phi_PII), 'mV')
print('Ratio =', (Phi_PII/RP_PII))

Phi_PII = 3290 mV
Ratio = 4.0
```

4.2 Complex Cyt -- Cytochrome bf

```
In [13]: disp.SVG('Cyt_abg.svg')
```

Out[13] :



```
In [14]: sbg.model('Cyt_abg.svg', convertR=True, convertCe=True, quiet=True)
         import Cyt_abg
```

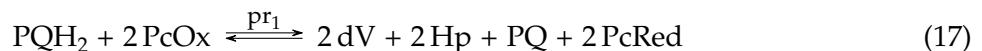
```
In [15]: ## Stoichiometry
sCyt = st.stoich(Cyt_abg.model(), quiet=True)
chemostats = ['PQ', 'PQH2', 'PcOx', 'PcRed', 'Hp', 'dV']
scCyt = st.statify(sCyt, chemostats=chemostats)
spCyt = st.path(sCyt, scCyt)
disp.Latex(st.sprintrl(sCyt, chemformula=True))
```

Out[15] :



```
In [16]: disp.Latex(st.sprintrl(spCyt, chemformula=True))
```

Out[16] :



```
In [17]: ## Compute net redox potential
RP_Cyt = (- redoxData.EpH('PQ/PQH2', pH=pH_p)
           - redoxData.VpH(pH_p - pH_n)
           + redoxData.E('PcOx/PcRed')
           )

print(RP_Cyt)
print('RP_Cyt = ', redoxData.mV(RP_Cyt), 'mV')
```

```
0.010757686938834443
```

```
RP_Cyt = 11 mV
```

```
In [18]: ## Compute the reaction potential Phi
phi = phiData.phi_species(phi_redox,spCyt['species'])
Phi_Cyt_ = -spCyt['N'].T@phi
Phi_Cyt = Phi_Cyt_[0][0]
print('Phi_Cyt =',redoxData.mV(Phi_Cyt), 'mV')
print('Ratio =', int(Phi_Cyt/RP_Cyt))
```

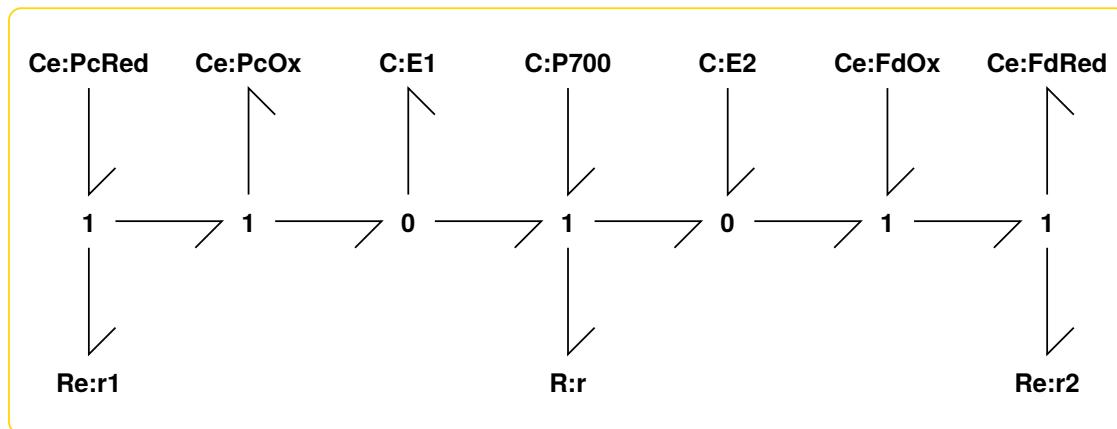
```
Phi_Cyt = 22 mV
```

```
Ratio = 2
```

4.3 Complex PI -- Photosystem I

```
In [19]: disp.SVG('PI_abg.svg')
```

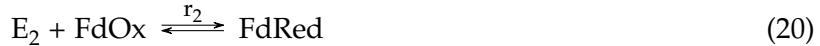
```
Out[19]:
```



```
In [20]: sbg.model('PI_abg.svg',convertR=True,convertCe=True,quiet=True)
import PI_abg
```

```
In [21]: ## Stoichiometry
sPI = st.stoich(PI_abg.model(),quiet=True)
chemostats = ['PcOx','PcRed','FdOx','FdRed','P700']
scPI = st.statify(sPI,chemostats=chemostats)
spPI = st.path(sPI,scPI)
disp.Latex(st.sprintrl(sPI,chemformula=True))
```

```
Out[21]:
```



In [22]: `disp.Latex(st.sprintrl(spPI,chemformula=True))`

Out [22]:



In [23]: `## Compute net redox potential`

```
RP_PI = (
    - redoxData.E('PcOx/PcRed')
    + V700
    + redoxData.E('FdOx/FdRed')
)

#print(RP_PI)
print('RP_PI =',redoxData.mV(RP_PI), 'mV')
```

`RP_PI = 961 mV`

In [24]: `## Compute the reaction potential Phi`

```
phi = phiData.phi_species(phi_redox,spPI['species'])
Phi_PI_ = -spPI['N'].T@phi
Phi_PI = Phi_PI_[0][0]
print('Phi_PI =',redoxData.mV(Phi_PI), 'mV')
print('Ratio =', int(round(Phi_PI/RP_PI)))
```

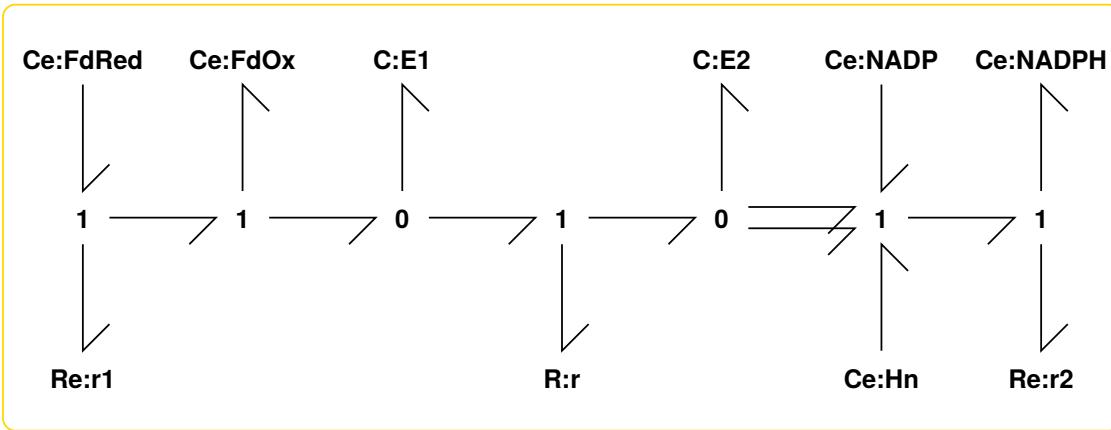
`Phi_PI = 961 mV`

`Ratio = 1`

4.4 Complex Fer -- Feredoxin-NADP reductase

In [25]: `disp.SVG('Fer_abg.svg')`

Out [25]:



```
In [26]: sbg.model('Fer_abg.svg', convertR=True, convertCe=True, quiet=True)
         import Fer_abg
```

```
In [27]: ## Stoichiometry
sFer = st.stoich(Fer_abg.model())
chemostats = ['FdRed', 'FdOx', 'NADP', 'NADPH', 'Hn']
scFer = st.stoich(Fer_abg.model(), chemostats=chemostats)
spFer = st.path(sFer, scFer)
disp.Latex(st.sprintrl(sFer, chemformula=True))
```

Swapping Re:r for two Sf in Fer
 Swapping Re:r1 for two Sf in Fer
 Swapping Re:r2 for two Sf in Fer
 Swapping Re:r for two Sf in Fer
 Swapping Re:r1 for two Sf in Fer
 Swapping Re:r2 for two Sf in Fer

Out [27] :



```
In [28]: disp.Latex(st.sprintrl(spFer, chemformula=True))
```

Out [28] :



```
In [29]: ## Compute net redox potential
RP_Fer = (
    - redoxData.E('FdOx/FdRed')
    + redoxData.EpH('NADP/NADPH', pH_n)
)

#print(RP_Fer)
print('RP_Fer =', int(1000*RP_Fer), 'mV')
```

RP_Fer = 105 mV

```
In [30]: ## Compute the reaction potential Phi
phi = phiData.phi_species(phi_redox, spFer['species'])
Phi_Fer_ = -spFer['N'].T@phi
Phi_Fer = Phi_Fer_[0][0]
print('Phi_Fer =', int(1000*Phi_Fer), 'mV')
print('Ratio =', int(Phi_Fer/RP_Fer))
```

Phi_Fer = 212 mV

Ratio = 2

5 The Electron Transport Chain

The overall model is described a bond graph tools file:

```
In [31]: ## File ETC_abg.py

import BondGraphTools as bgt
import PII_abg
import Cyt_abg
import PI_abg
import Fer_abg

def model():
    """
    Model of chloroplast electron transport chain
    """

    ETC = bgt.new(name='ETC')      # Create system
    PII = PII_abg.model()
    Cyt = Cyt_abg.model()
    PI = PI_abg.model()
    Fer = Fer_abg.model()
    bgt.add(ETC, PII, Cyt, PI, Fer)

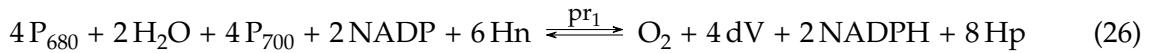
    return ETC
```

5.1 Unify species in model using mbg.unify()

```
In [32]: import ETC_abg
model = ETC_abg.model()
common = ['PQ', 'PQH2', 'PcOx', 'PcRed', 'FdOx', 'FdRed', 'Hn', 'Hp']
mbg.unify(model, common, quiet=True)
s = st.stoich(model, quiet=True)

In [33]: chemostats = ['H2O', 'O2', 'NADP', 'NADPH', 'Hp', 'Hn', 'P680', 'P700', 'dV']
sc = st.statify(s, chemostats=chemostats)
sp = st.path(s, sc)
disp.Latex(st.sprintrl(sp, chemformula=True))
```

Out[33] :



```
In [34]: ## Compute the reaction potential Phi
phi = phiData.phi_species(phi_redox, sp['species'])
Phi_ = -sp['N'].T@phi
Phi = Phi_[0][0]
print('Phi =', redoxData.mV(Phi), 'mV')
```

Phi = 7603 mV

References

Robert E. Blankenship. *Molecular mechanisms of photosynthesis*. Wiley Blackwell, Oxford, 2015.

Robert E. Blankenship and Roger C. Prince. Excited-state redox potentials and the z scheme of photosynthesis. *Trends in Biochemical Sciences*, 10(10):382 – 383, 1985. ISSN 0968-0004. doi:[10.1016/0968-0004\(85\)90059-3](https://doi.org/10.1016/0968-0004(85)90059-3).

David G Nicholls and Stuart Ferguson. *Bioenergetics 4*. Academic Press, Amsterdam, 2013.